

Policy: SC-20-103
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Title: Pest Control Procedures

I. Purpose:

The purpose of this policy is to outline expectations for prevention and control the entrance of pests and eradication of infestations in facilities housing research and teaching animals. The Animal Welfare Act and Regulations (5/1/2023 Edition) and the Guide for the Care and Use of Laboratory Animals (The Guide, Eighth Edition), require a pest control program as a component of the animal care program.

II. Policy:

All units housing laboratory or teaching animals must have standard operating procedures on pest control. Refer to the Policy and Procedures Manual for pest control guidance <https://ucdavispolicy.ellucid.com/documents/view/295>.

III. Procedure:

Building design, construction, and maintenance are the primary components to preventing entrance of potential pests into animal areas. All units must ensure that windows, doors, and exterior walls are sealed to prevent entrance of pests and predators. Interior walls, drains, and vents must be checked for cracks and leaks and repaired as needed. Local facilities management groups must be contacted when repairs are needed that require their services.

Facilities shall be kept free of clutter, unnecessary storage of equipment, and trash to prevent the harborage of pests. Keep cardboard boxes, feed bags, etc. off of the floor. Refer to the [Standard of Care SC-50-101 Housekeeping in Facilities housing Biomedical Research animals](#) and [the Standard of Care SC-50-106 Housekeeping in Facilities Housing Agricultural Animals](#) for further clarification.

Animal units are required to develop pest control standard operating procedures specific to their pest control needs. A regularly scheduled and documented program of control and monitoring should be implemented (*Guide, p.74*).

Facility managers or designee must coordinate pest control procedures with Facilities Management as needed to minimize harmful effects to the integrity of building and equipment construction as well as to ensure the safety of students and staff.

Principal Investigators and research staff must be consulted before ANY pesticides or other substances will be used in animal areas or around caging, food or other items that will contact the animals to prevent any adverse effects or unintended consequences that could affect the research experiments.

When necessary for the prevention or control of potential risks associated with pests, nontoxic substances and live traps can be utilized.

Any traps used should be humane (*Guide, p74*). All live and lethal (e.g., snap) traps must be checked and animals removed daily (*Guide, p.74*). Alternatives to "sticky/adhesive" live board traps should be used for mice whenever possible to avoid unnecessary animal distress as required by the Guide as part of a program of adequate veterinary care (AAALAC Position Statement, Frequency of Monitoring Rodent Traps) (<https://www.aaalac.org/accreditation-program/faqs/#D8>).

Proper protective equipment MUST be worn when handling wild rodents. Minimally disposable gloves, an outer covering (coat or uniform), and a barrier mask (surgical mask to prevent splash exposure, N95 however, is recommended) are required for handling live wild rodents.

The pans under primary enclosures with grill-type floors and the ground areas under raised runs with mesh or slatted floors must be cleaned as often as necessary to prevent accumulation of feces and food waste and to reduce disease, hazards, pests, insects, and odors (AWA, p. 148). The same procedure applies to animal holding areas of terminal facilities (AWA, p. 228). Pens, runs, and outdoor housing areas using material that cannot be sanitized using live steam under pressure or by washing with hot water, soap or detergent such as gravel, sand, grass, earth, or absorbent bedding must be sanitized by removing the contaminated material as necessary to prevent the aforementioned hazards (AWA, p. 149)

Zoonotic agents have been identified in the wild rodent populations in Davis. Live trapped rodents found by Animal Care Staff MAY be submitted for surveillance to the Comparative Pathology Laboratory (CPL). CPL must be informed that the animals are wild caught, so they can take proper safety precautions. Transport is best accomplished by placing the entire live trap in a clean rodent cage with filter top. The filter top lid should be taped on the cage. The cage is put in the euthanasia chamber, and euthanasia performed, before handling the wild rodent. This submission or euthanasia must be completed on the day the animal is discovered by the Animal Care Staff in the live trap. Submission of animals to CPL must be coordinated with the Campus Veterinary Services at LAHC@ucdavis.edu or 530-752-0514.

Use of commercial or regulated pesticides and aerosolized products requiring special applicators such as a back pack should be done in consultation with Facilities Management and should only be used when other preventive or control methods are unsuccessful. These Pesticides when used indoors must be documented. Principal Investigators must also be aware of and agree on any commercial or regulated pesticides used in animal areas or around caging, food or other items that will contact the animals.

Facility managers or designees must be appropriately trained on pest control procedures and proper implementation of pest control methods.